

Physiological and pathological functions of Reactive Nitrogen Species (RNS) and Reactive Sulphur Species (RSS) on Male Reproductive functions

Sulagna Dutta^{1*}, Pallav Sengupta^{2*}, Antony V. Samrot³

¹School of Life Sciences, Manipal Academy of Higher Education (MAHE), Dubai 345050, UAE. ²Department of Biomedical Sciences, College of Medicine, Gulf Medical University, Ajman, UAE. ³Faculty of Medicine, Biosciences and Nursing, MAHSA University, Malaysia.

Received on: 21-Jul-2023, Accepted and Published on: 07-Nov-2023

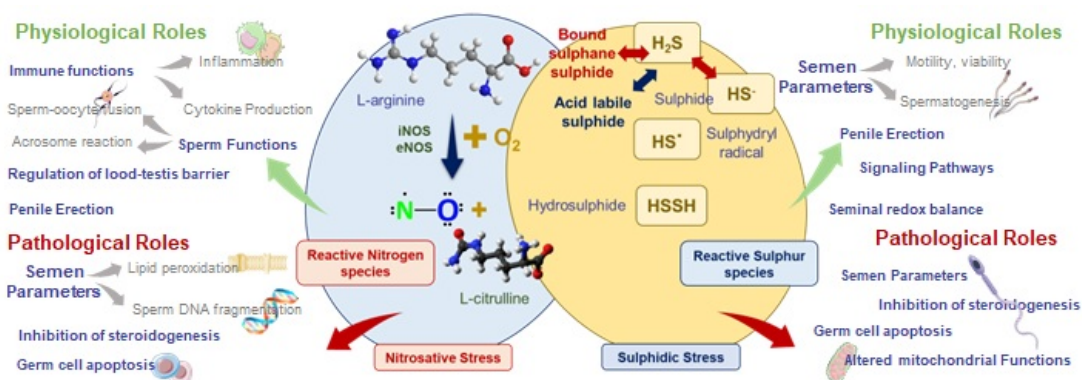
Review

ABSTRACT

Reactive Nitrogen Species (RNS) and Reactive Sulphur Species (RSS) play crucial roles in numerous physiological pathways, including cellular signaling, metabolic cascades, and redox balance maintenance. In male reproduction, these molecules serve dual purposes: they support

essential physiological functions but can cause harm if unregulated. This review explores the roles of RNS and RSS in male reproduction, emphasizing their beneficial and detrimental impacts. Physiologically, RNS and RSS aid in sperm maturation, capacitation, and initiating the acrosome reaction. Regulated synthesis of these species is vital for redox-regulated events essential for optimal male reproductive outcomes. Specifically, nitric oxide, a primary RNS, regulates sperm functions and penile erection mechanisms. However, imbalances, leading to excessive RNS and RSS levels, can cause oxidative and nitrosative stresses, which can trigger lipid peroxidation, protein alterations, and sperm DNA fragmentation, affecting sperm vitality, motility, and genomic integrity.

Keywords: male infertility; nitrosative stress; reactive nitrogen species; reactive oxygen species; reactive sulphur species



INTRODUCTION

Reactive species, especially those derived from oxygen, have been notably studied in biomedicine due to their ambivalent influence in both physiological and pathological contexts.¹⁻³ Conversely, the roles of Reactive Nitrogen Species (RNS) and Reactive Sulphur Species (RSS) are relatively understudied, especially concerning male reproductive functions. RNS and RSS are groups of molecules that originate from their primary nitrogen

and sulphur precursors, respectively. These species are integral to various biological processes, functioning as signaling agents that orchestrate cellular activities.^{4,5}

Within the male reproductive framework, physiological concentrations of RNS and RSS are pivotal for a spectrum of processes from spermatogenesis to influencing sperm motility and efficacy.^{5,6} For example, nitric oxide (NO), an essential RNS constituent, plays a central role in penile erection and sperm functionality, underscoring its importance in male fertility.⁷ In parallel, RSS, while a more nascent research area, holds potential to reveal novel physiological roles pertaining to male reproduction.

Conversely, an excessive accumulation of these species, stemming from external influences like environmental contaminants or inherent causes such as oxidative stress, can be injurious.⁸ Heightened RNS and RSS levels may culminate in nitrosative or sulphidic stress, respectively.^{4,9} These stressors are associated with multiple maladies in the male reproductive system,

*Corresponding Author: Dr. Sulagna Dutta, Dr. Pallav Sengupta
Email: sulagna_dutta11@yahoo.com (SD), pallav_cu@yahoo.com (PS)

Cite as: *J. Integr. Sci. Technol.*, 2024, 12(3), 755.
URN:NBN:sciencein.jist.2024.v12.755



©Authors CC4-NC-ND, ScienceIN ISSN: 2321-4635
http://pubs.thesciencein.org/jist

encompassing diminished sperm quality, compromised sperm movement, and even instances of male infertility.⁹

The objective of this literature review is to provide an exhaustive examination of the physiological and pathological connotations of RNS and RSS within male reproductive mechanisms. By elucidating both their salutary and deleterious impacts, this review seeks to address present knowledge deficits and foster forthcoming research directions. Such endeavors could conceivably enhance diagnostic and therapeutic methodologies pertaining to male reproductive wellbeing.

REACTIVE NITROGEN SPECIES

Reactive nitrogen species (RNS) constitute a diverse group of molecules, each exhibiting unique chemical characteristics and reactivity, all originating from a shared precursor: $\cdot\text{NO}$. This $\cdot\text{NO}$ is recognized as a free radical and a mild oxidizing agent, playing a pivotal role in regulating critical cellular processes across a spectrum of organ systems, encompassing the cardiovascular, neural, immune, reproductive, gastrointestinal, and secretory systems.^{10,11} $\cdot\text{NO}$ synthesis predominantly arises from an enzymatic process mediated by nitric oxide synthase (NOS). There exist three distinct isoforms of NOS: endothelial NOS (eNOS), inducible NOS (iNOS), and neuronal NOS (nNOS).¹⁰ These isoforms vary in their affinities for calmodulin and the magnitude of $\cdot\text{NO}$ production. Intracellularly, $\cdot\text{NO}$ can manifest dual roles, ranging from

physiological to pathological outcomes. The specific effects imparted by $\cdot\text{NO}$ are contingent upon its concentration, which is intrinsically linked to its rate of synthesis and the prevailing cellular redox environment.^{10,12} At nanomolar concentrations, $\cdot\text{NO}$ operates through the traditional cGMP-dependent signaling pathway, which entails the activation of soluble guanylyl cyclase (sGC), cGMP generation, followed by the stimulation of particular cGMP-dependent enzymatic targets.¹³ In contrast, at elevated, micromolar concentrations, $\cdot\text{NO}$ employs non-traditional, indirect signaling mechanisms independent of cGMP.^{14, 15} This mode of action involves covalent post-translational modifications of proteins, notably S-nitrosylation, S-glutathionylation, and tyrosine nitration. Such non-traditional signaling arises from $\cdot\text{NO}$ interactions with prevalent intracellular reactants like O_2 or $\text{O}_2\cdot^-$, leading to the generation of various RNS (like nitrogen dioxide, $\cdot\text{NO}_2$; peroxynitrite, ONOO^- ; nitroxyl, HNO ; dinitrogen trioxide, N_2O_3 ; and dinitrogen tetroxide, N_2O_4). These RNS exhibit higher reactivity compared to $\cdot\text{NO}$ and have the capability to modify cysteine and tyrosine residues within proteins.^{16,17} Notably, S-nitrosylation and S-glutathionylation are dynamic processes, and their equilibrium can be modulated by specific reductases, namely thioredoxin and S-nitrosoglutathione reductase.^{18,19} Conversely, tyrosine nitration, predominantly facilitated by ONOO^- , is largely an irreversible process and can often lead to cytotoxic implications of $\cdot\text{NO}$ by potentially altering key protein functions.^{19,20}

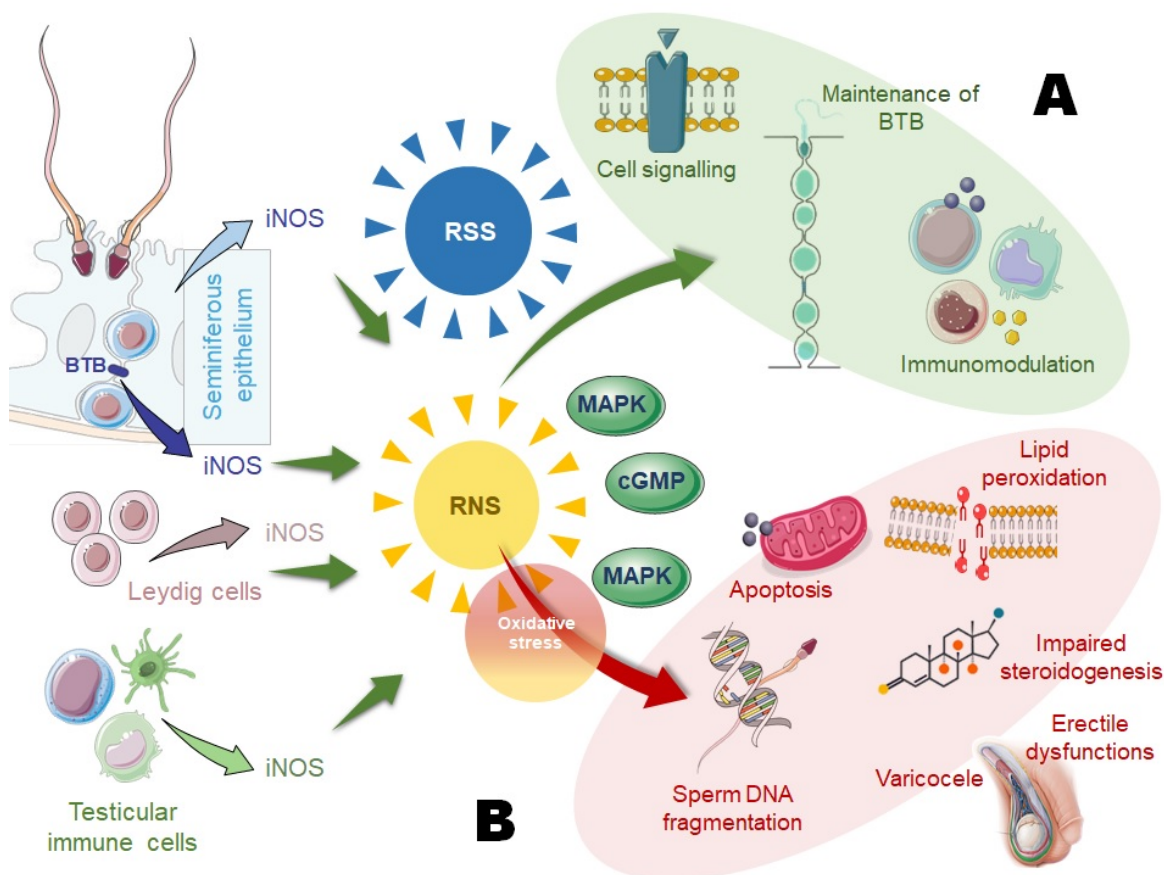


Figure 1. Physiological (A) and pathological roles of reactive nitrogen species (RNS) and reactive sulphur species (RSS) on male reproduction.

Sources of RNS in male reproductive system

In the human reproductive system, NO synthesis pathway plays a pervasive role, with NO modulating numerous reproductive processes.^{21, 22} Within the testicular tissue, all three NOS isoforms are present across various cell categories—namely Leydig, endothelial, myoid, Sertoli, and germ cells situated in the seminiferous epithelium.^{23, 24} In penile tissue, the neuronal form of NOS (nNOS) is identified in the pelvic plexus, dorsal penile and cavernous nerve, and within the smooth muscle cells of the corpus cavernosum.^{25, 26} In contrast, endothelial NOS (eNOS) is detected in both cavernous endothelial cells and corpus cavernosum smooth muscle cells.²⁶ Human sperm cells synthesize NO through the function of all known NOS isoforms, which are localized in either the sperm head or flagellum.^{21, 22} Notably, as sperm undergo maturation, there is a variability in the enzymatic activity of sperm-associated NOS, highlighting the intrinsic physiological significance of NO in sperm functions (Fig. 1).²⁷

Cellular signaling of RNS in male reproductive events

RNS are instrumental in modulating diverse cellular activities. Within this category, NO stands out as a crucial mediator in male reproductive biology. This composition explores the complex signaling pathways of NO related to male reproduction, with a particular focus on its integral involvement in sperm activity and male reproductive tissue dynamics. NO , produced within the reproductive system, is a multifunctional entity that performs numerous roles. At physiological levels, NO is generated by Leydig cells, facilitating unimpeded diffusion across cell boundaries, thus enhancing testicular functionalities. Its regulatory scope over the male reproductive apparatus spans processes like spermatogenesis, sperm maturation, motility, and germ cell programmed cell death.^{23, 25} The predominant mechanism through which NO enforces these controls is primarily by triggering the soluble guanylate cyclase (sGC) and subsequently signaling through the cyclic guanosine monophosphate (cGMP) protein kinase G (PKG) route.²⁴ Moreover, penile tissues significantly benefit from NO 's actions, where it plays a foundational role in the erectile mechanism. This functional attribute is rooted in the generation of NO by constitutive NOS, especially eNOS and nNOS.^{28, 29} Interference with these NOS variants, through genetic or drug-based means, highlights the indispensable nature of NO in mediating erectile capabilities.^{30, 31} Within this scope, NO 's regulatory authority primarily hinges on the sGC/cGMP signaling framework.³²

Concerning spermatozoa, NO is a proactive modulator of numerous molecular pathways pivotal for assorted sperm operations. Such operations encompass sperm motility, survival, hyperactivation, capacitation, acrosome reaction (AR), oocyte fusion, and the subsequent programmed cell death, all pivotal phases for effective fertilization and procreation.^{21, 22, 33} While the sGC/cGMP pathway remains recurrent for many of these roles, an essential mechanism merits attention: the tyrosine nitration of distinct sperm proteins. This nitration has dual implications in sperm biology, with research indicating both detrimental.^{34, 35} and advantageous^{21, 36} outcomes, implying a sophisticated and harmonized regulation. Beyond these roles, NO has been proven to serve a defensive role against oxidative stress in spermatozoa. It

offers protection to sperm cells against lipid peroxidation, predominantly by safeguarding protein sulfhydryl clusters, thereby averting potential oxidative membrane damage.^{37, 38} Thus, as an integral component of the RNS group, NO wields profound effects on diverse facets of male reproductive phenomena. From its deep-rooted cellular signaling in sperm cells to its cardinal role in erectile dynamics, the ubiquitous nature of NO within male reproductive biology is evident, underscoring the need for deeper research for potential therapeutic prospects.

Sperm motility and viability

RNS, notably NO , are integral to the complex biochemical mechanisms that govern sperm movement and survival. Throughout the intricate physiological journey within the reproductive system, NO 's involvement becomes progressively vital, safeguarding the prime functioning of sperm imperative for effective fertilization. Numerous studies have highlighted the crucial modulatory function of NO in dictating sperm motility. Research that pharmacologically adjusted NO concentration reinforced this interrelation.^{39, 40} For instance, the application of sodium nitroprusside (SNP), a recognized NO donor, at concentrations between 25-100 nM, beneficially supported the maintenance of post-thaw human sperm activity and survival.⁴⁰ Conversely, introducing NG-nitro-L-arginine methyl ester (L-NAME) to the sperm culture medium resulted in a marked decrease in human sperm motility.^{41, 42} Such outcomes accentuate the fundamental role of NO in steering sperm kinetics. A noteworthy revelation was the reduced nitrite production in asthenozoospermic individuals compared to normozoospermic ones, emphasizing the crucial nature of intrinsic NO not only in modulating sperm movement but also its fertilization capability.^{43, 44} On a molecular level, NO modulates sperm motility by triggering soluble guanylate cyclase (sGC), which in turn activates cGMP-dependent protein kinases. Recent insights reveal the function of NO in amplifying sperm movement through the bolstering of energy generation within sperm mitochondria.⁴⁵ Specifically, a 3-hour exposure of spermatozoa in Tyrode's medium led to reductions in sperm movement, mitochondrial membrane potential, NO content, and eNOS expression. Yet, the incorporation of M40403, a superoxide dismutase (SOD) analogue, prominently rejuvenated eNOS expression, raising NO concentrations. The operational state of sperm mitochondria, indicated by the proportion of positive MitoTracker@Green FM sperm cells, increased significantly in sperm samples with enhanced NO concentration. Additionally, a rise in NO concentration in semen samples favorably affected another dimension of sperm mitochondrial functionality. This related to heightened mRNA expression of the nuclear-encoded subunits of both complex I and IV of the Electron Transport Chain (ETC), with subunits dictated by the mitochondrial DNA returning to baseline values. Such findings correspond positively with sperm movement, a chief indicator of semen quality, clarifying that NO improves the sperm's capability to fertilize.^{46, 47} These results also unveiled, unprecedentedly, the effectiveness of redox-active compounds in enhancing sperm fertilization potential during sperm extraction and preparation for in vitro fertilization. Beyond movement and energy production, NO is pivotal for upholding standard sperm morphology, an essential metric for the precise

prediction of fertility potential and in vitro fertilization pregnancy outcomes. Thus, the diverse functionalities of $\cdot\text{NO}$ and RNS in sperm physiology underline their significance in reproductive medicine, emphasizing the imperative for ongoing research in this domain.⁴⁸

Sperm capacitation

Sperm capacitation represents an essential physiological transformation that equips spermatozoa with the capability to fertilize an ovum. This metamorphosis is delineated by a multitude of biophysical and biochemical modifications.⁴⁹ Contemporary research has illuminated the pivotal function of RNS, particularly $\cdot\text{NO}$, in orchestrating this complex procedure (**Fig. 1**). When human spermatozoa were exposed to minor concentrations of $\cdot\text{NO}$ -releasing agents, it elucidated the importance of $\cdot\text{NO}$ in capacitation.⁴⁹ Such experimentation underscored that $\cdot\text{NO}$ donors augment capacitation, while in contrast, NOS antagonists considerably diminish this process.⁵⁰ These outcomes authenticate the essential influence of intrinsic $\cdot\text{NO}$ in empowering spermatozoa to reach optimal fertilizing capability. Concurrently, this research unveiled a consistent $\cdot\text{NO}$ -driven modulation of capacitation, regardless of the specific stimulus, whether albumin, progesterone, or fetal cord serum ultrafiltrate.^{51, 52}

Examining the molecular details, it was ascertained that $\cdot\text{NO}$ synthesis in spermatozoa, which commences at the beginning of capacitation, presides over several pivotal biochemical phenomena. Such phenomena encompass the amplification in cyclic AMP (cAMP) concentrations and the comprehensive serine/threonine phosphorylation of protein kinase A (PKA) targets. In addition, $\cdot\text{NO}$ adjusts the tyrosine phosphorylation of fibrous sheath constituents and concurrently induces tyrosine nitration in assorted proteins.²² Notably, these $\cdot\text{NO}$ -driven changes persist throughout the capacitation procedure, emphasizing its essential role in preparing the sperm for efficacious fertilization. Another dimension to the $\cdot\text{NO}$ -mediated mechanism in capacitation pertains to the genesis of its synthesis. Evidence suggests the involvement of multiple NOS isoforms in capacitation, hinting at a coordinated effort of several NOS variants to regulate $\cdot\text{NO}$ concentrations and consequently influence the capacitation trajectory.⁵³ Thus, the revelations regarding the influence of $\cdot\text{NO}$, an integral component of the RNS category, have revolutionized our perspective on sperm capacitation. Its role as a chief regulatory entity, directing spermatozoa through requisite biochemical transformations, and its continuous involvement during capacitation, accentuates its vital character.⁵ The diverse effects of $\cdot\text{NO}$, from influencing cAMP concentrations to managing phosphorylation episodes, firmly establish it as a key molecule in ensuring the reproductive efficacy of spermatozoa. As the molecular dynamics of sperm capacitation continue to be elucidated, the prominence of $\cdot\text{NO}$ and the larger RNS family is poised to persistently dominate discussions in reproductive biology investigations.^{54, 55}

Hyperactivation

RNS encompass a collection of highly active free radicals that originate from nitrogen. In the field of reproductive biology, these molecular entities have attracted substantial consideration, especially in decoding the complex mechanisms that underpin sperm activity.⁵ An essential function of sperm is hyperactivation,

a distinct and intense motility pattern marked by irregular flagellar oscillations, which is crucial for the ability of the sperm to fertilize.⁵ The influence of RNS, particularly $\cdot\text{NO}$, in modulating sperm hyperactivation has become a prime focus of contemporary scientific investigation.⁷ Traditionally, the association between $\cdot\text{NO}$ and sperm motility was underscored, with a significant differentiation made depending on the $\cdot\text{NO}$ concentration. $\cdot\text{NO}$ exerts bifunctional effects on sperm hyperactivation contingent on its concentration.^{41, 56} At concentrations below 1 μM , $\cdot\text{NO}$ appears to enhance sperm hyperactivation. This might be a result of various molecular processes such as increased intracellular calcium, alteration in adenylate cyclase activity, or possible activation of specific protein kinases. These processes potentially impact axonemal proteins and dynein arms, which dictate the propulsive force and specialized oscillatory patterns of hyperactivated sperm. In contrast, when $\cdot\text{NO}$ concentrations exceed 1 μM , a decline in hyperactivated sperm motility is documented.⁴² Elevated $\cdot\text{NO}$ levels might induce oxidative stress, potentially causing lipid peroxidation in the sperm membrane and subsequent reduction in membrane fluidity.⁵⁷ Such disturbances might negatively impact cellular signaling mechanisms that sustain hyperactivation. Moreover, excessive $\cdot\text{NO}$ could hinder mitochondrial operations, jeopardizing the ATP generation essential for the energy-demanding hyperactivated movement.⁴¹ It is noteworthy that the influence of $\cdot\text{NO}$ on sperm hyperactivation is analogous to its effect on standard sperm motility, as highlighted in various research works.^{42, 58} This emphasizes the critical equilibrium sperm cells must preserve in their intracellular milieu to function optimally. Clearly, an intricate balance of $\cdot\text{NO}$ is vital, serving almost as a molecular regulator, either enhancing or suppressing hyperactivated movement based on its concentration. Nonetheless, it's imperative to recognize that while $\cdot\text{NO}$ is a decisive factor, sperm motility, including hyperactivation, arises from a complex interplay of numerous molecular interactions and signaling pathways.^{42, 59} RNS represent just one component in this sophisticated orchestration. Future studies should center on elucidating the wider interactions of RNS with other reactive entities and their collective impacts on the diverse facets of sperm activity. Thus, RNS, especially $\cdot\text{NO}$, are crucial in shaping sperm hyperactivation.⁴² This revelation not only offers a more profound understanding of the intricate mechanisms of sperm activity but also paves the way for potential therapeutic strategies in addressing male infertility issues.^{43, 45, 46}

Acrosome reaction

RNS are integral to a myriad of physiological functions, with a pronounced importance in reproductive biology, especially concerning the acrosome reaction (AR). The AR represents a critical phase during fertilization, wherein the acrosomal membrane of sperm merges with the adjacent plasma membrane, leading to the discharge of enzymes crucial for penetrating the zona pellucida (ZP) and facilitating sperm-egg fusion. Among the RNS group, $\cdot\text{NO}$ stands out, having been associated with modulating ZP adhesion and the AR. Multiple pieces of evidence reinforce the relevance of $\cdot\text{NO}$ in this arena.²¹ A prominent research finding indicates that enriching the capacitation medium of human sperm with modest amounts of SNP augments the spermatozoa adhering

to hemizona.⁶⁰ This hints at direct impact of $\cdot\text{NO}$ on fortifying sperm-ZP adhesion. Additionally, this enrichment seems to bolster sperm's affinity toward the ZP, underscoring $\cdot\text{NO}$'s potential role as an enhancer in this mechanism. On the contrary, the introduction of L-NAME, a NO synthesis inhibitor, to capacitated sperm led to a significant decline in the fusion capability.⁶¹ Noteworthy, this impact did not encompass ZP adhesion, implying that while $\cdot\text{NO}$ is crucial for the fusion mechanism, its involvement in ZP adhesion is more complex.⁶¹ Additionally, tests employing $\cdot\text{NO}$ -releasing agents have revealed their capacity to trigger the AR in human spermatozoa in a dose-responsive manner.⁶⁰ These inductions, when compared with their respective controls, exhibit pronounced distinctions, emphasizing $\cdot\text{NO}$'s indispensable role in the AR.^{48, 50} In contrast, hemoglobin, serving as a $\cdot\text{NO}$ neutralizer, has demonstrated the capability to hinder the AR in spermatozoa pre-treated with follicular fluid. This observation bolsters the foundational role $\cdot\text{NO}$ assumes in modulating the acrosome reaction.⁶² Thus, the nuanced interactions between RNS, especially $\cdot\text{NO}$, and the mechanisms of ZP adhesion and acrosome reaction illuminate the intricate chemical communication inherent in human reproductive processes. Grasping these molecular pathways presents potential therapeutic breakthroughs in reproductive medicine.^{48, 63}

Changes in sperm redox status change gene transcription?

Sperm cells, specifically spermatozoa, have been traditionally understood as transcriptionally inactive due to their highly compacted chromatin structure. This has been attributed to the replacement of histones with protamines in mature sperm, which was thought to suppress transcriptional activity. However, recent studies have indicated that approximately 15% of the sperm nuclear material still maintains a connection with histones, which adopt a characteristic nucleosomal configuration.⁶⁴ This preservation implies a potential for transcriptional processes in these nucleosomal areas, particularly since they show specific associations with transcription factors and RNA polymerase. Recent studies emphasize the significance of this frequently disregarded nucleosomal section in male reproductive cells. These discoveries signify a reevaluation of sperm cell biology, suggesting these regions might be more critical than previously believed. A pivotal observation was the association between RNS, especially $\cdot\text{NO}$, and gene transcription in sperm cells. Studies on this relationship have elucidated the connection between the redox state and gene expression modulation in spermatozoa. Specifically, sperm samples with increased $\cdot\text{NO}$ levels displayed enhanced mRNA expression of antioxidative enzymes, such as catalase and MnSOD.⁴⁶ This implies that $\cdot\text{NO}$ could enhance the antioxidative response in sperm by promoting the transcription of catalase and MnSOD.²² Additionally, alterations in the redox equilibrium of spermatozoa seem to have broader consequences for gene transcription. This is supported by the documented upregulation of nucleus-encoded components of both complex I and IV of the Electron Transport Chain (ETC) in sperm samples with heightened $\cdot\text{NO}$ levels.⁶² The relationship between $\cdot\text{NO}$ and gene transcription modulation emphasizes the interrelation between RNS and the sophisticated redox system in sperm cells, pointing to new areas of exploration in male fertility research.⁶⁵

Pathophysiological roles of RNS on male reproduction

RNS are integral to numerous physiological and pathological mechanisms within the organism. $\cdot\text{NO}$, a key constituent of the RNS group, possesses a dual functionality of significant interest, especially concerning male reproductive functions. The interaction of $\cdot\text{NO}$ with fertility and sperm activity is multifaceted and depends on its concentration, making it a multifunctional entity in reproductive biology. The dualistic action of $\cdot\text{NO}$ is not limited to reproductive systems but is observed in multiple physiological systems. At physiological, lower concentrations, $\cdot\text{NO}$ facilitates essential events related to fertilization. Conversely, at augmented levels, its impacts are harmful. This bidirectional effect has led to inconsistencies in scientific findings about $\cdot\text{NO}$'s exact relationship with fertility and sperm performance.^{22, 29}

The theory suggesting $\cdot\text{NO}$ toxicity proposes that excessive quantities of this compound can inhibit mitochondrial respiration, reduce ATP reserves, and initiate DNA oxidative, nitration, and deamination processes.^{37, 66} Research has documented reduced sperm motility in the context of elevated pure $\cdot\text{NO}$ concentrations and other $\cdot\text{NO}$ sources like SNP or S-nitroso-N-acetylpenicillamine. Such results are consistent with clinical data showing a relationship between high $\cdot\text{NO}$ levels in semen and infertility or diminished sperm motility.^{37, 47, 66, 67} Additionally, an increase in $\cdot\text{NO}$ concentration in sperm cells from infertile individuals corresponds with a decline in sperm metabolism.^{5, 68} Both sperm viability and structural integrity are negatively influenced by high $\cdot\text{NO}$ concentrations.^{68, 69} For instance, specific SNP concentrations have been shown to significantly decrease sperm viability, while elevated $\cdot\text{NO}$ levels result in structural anomalies in sperm.⁷⁰ Furthermore, the effects of excess $\cdot\text{NO}$ aren't restricted to sperm. In testicular tissues, increased $\cdot\text{NO}$, followed by peroxynitrite formation, curtails testosterone release. Excessive production can also lead to blood vessel vasodilation and extended myofibroblast relaxation, which might obstruct essential sperm transport movements.^{59, 71} Within Sertoli cells, surplus $\cdot\text{NO}$ can modify the patterns of actin-associated proteins crucial for cellular processes like adhesion, proliferation, migration, and differentiation.^{72, 73} In the context of penile tissue, elevated $\cdot\text{NO}$ concentrations, particularly from inducible NOS (iNOS), can instigate pathophysiological alterations.⁵⁵

While the negative implications of augmented $\cdot\text{NO}$ levels are clear, reproductive mechanisms also display vulnerability to decreased $\cdot\text{NO}$ concentrations.⁷⁴ Vasculogenic erectile disorders, for instance, have been linked to lowered NOS activity, indicating potential erectile issues due to decreased $\cdot\text{NO}$ availability. Seminal plasma concentrations of $\cdot\text{NO}$ and 3-nitrotyrosine (3-NT) are critical for sperm performance. Discrepancies in these concentrations in infertile individuals' seminal plasma have been documented, although the exact nature of these variations remains debated.^{75, 76} Increased 3-NT concentrations in seminal plasma from men with idiopathic infertility hint at the role of intensified oxidative stress in reduced reproductive capacity.^{35, 77} Some research, such as that conducted by Huang and colleagues⁷⁶, suggests that even marginally reduced $\cdot\text{NO}$ levels in seminal plasma could increase infertility risk, irrespective of statistical significance. The complex role of $\cdot\text{NO}$ in male reproductive

biology emphasizes its significance in ensuring ideal sperm functionality. Both excessively low and high NO concentrations can negatively impact sperm count, movement, and structure, potentially leading to infertility. This nuanced balance accentuates the urgency for a more comprehensive understanding the role of RNS in reproduction, offering potential avenues for advanced therapeutic strategies aimed at addressing male infertility.^{5,78}

REACTIVE SULFUR SPECIES

RSS is characterized as redox-reactive sulfur-bearing entities capable of either oxidizing or reducing biological macromolecules under standard physiological conditions.⁷⁹ Analogous to ROS and RNS, RSS originate intracellularly and have gained recognition as pivotal biomolecular regulators in the biological domain, playing roles in redox-mediated signal transduction by modulating cysteine-based redox alterations in proteins, genomic modulation, ion flux, mitochondrial operations, metabolic activities, among other functions.⁶⁵ Notable members of these entities encompass thiol radicals (RSS), disulfide-S-oxides [RS(O)2SR], and sulfenic acids (RSOH). Additionally, H₂S, representing a fully reduced sulfur variant, qualifies as an RSS due to its ability to undergo redox interactions with proteins, influencing their functional activities.⁴ The chemical biology inherent to these diverse sulfur redox states governs their characteristics and modus operandi. This discourse places an emphasis on H₂S, given its intriguing implications across a spectrum of biological platforms realized through a variety of operational mechanisms.^{80, 81} Naturally, H₂S formation within mammalian cellular structures results from L-cysteine processing, facilitated by two pyridoxal-50-phosphate reliant enzymes: cystathionine-β-synthase (CBS) and cystathionine-γ-lyase (CSE). Beyond L-cysteine, CBS has the capability to utilize homocysteine for cystathionine synthesis, which subsequently, under CSE's influence, gives rise to H₂S. Another H₂S biosynthesis pathway involves the pyridoxal-50-phosphate independent enzyme, 3-mercaptopyruvate sulfurtransferase (3-MST), in conjunction with cysteine aminotransferase.⁸¹ Intrinsic H₂S formation also occurs through non-catalytic routes and via elemental sulfur derivatives, such as thiosulfate and thiocysteine, which can be reduced to H₂S within the glycolytic cascade.⁷⁹ Post-biosynthesis, H₂S partakes in reactions with an assortment of biological entities like ROS/RNS, cysteine-related compounds, and metalloproteins.^{80, 81} Such interactions elucidate the biological roles attributed to H₂S.

Synthesis of RSS in male reproductive system

Hydrogen sulfide (H₂S) within the reproductive system has historically been recognized for its cytotoxic effects and negative correlations with fecundity.⁸² Yet, recent investigations indicate that H₂S is intrinsically synthesized and plays a pivotal role in orchestrating a multitude of reproductive functions across both sexes.^{83, 84} The enzymes responsible for H₂S biosynthesis have been identified in diverse sections of the male reproductive anatomy. In human testicular tissues, immunohistochemical analyses have demarcated a specific locational distribution of two key H₂S biosynthetic enzymes: CBS is localized in Leydig, Sertoli, and germinal cells, whereas CSE is predominantly observed in Sertoli cells and progenitor germinal cells like spermatogonia.⁸³ This particular spatial segregation of CSE and CBS is similarly

discerned in human penile tissues. To elucidate further, both CBS and CSE are present within the trabecular muscular structures. Additionally, CSE is distinctly present in the vascular smooth muscle cells and peripheral neural structures, with their enzymatic activity being manifested in homogenates derived from penile tissue.⁸⁵

Physiological Signaling of RSS in Male Reproductive Events

Reactive sulfur species (RSS) are integral to many physiological processes, including those related to the male reproductive system. Hydrogen sulfide (H₂S) stands out as a crucial RSS in regulating male reproductive activities. The physiological significance of H₂S in male reproductive events includes its roles in erectile function, spermatogenesis, and sperm motility, underlining its importance in male fertility. The association between H₂S and the male reproductive system was initially evidenced through studies using sodium hydrosulfide (NaHS), an H₂S donor. In primates, NaHS demonstrated the beneficial impact of H₂S on erectile functionality.⁸⁶ Subsequent research further corroborated this, showing that exogenous application of H₂S to human corpus cavernosal tissues improved erectile response.⁸⁷ Conversely, when the internal generation of H₂S was obstructed using CSE and CBS inhibitors, a significant reduction in erectile functionality was observed. This highlights the essential role of internally generated H₂S in erectile function and suggests potential therapeutic applications of H₂S modulation for addressing erectile dysfunction in males.⁸⁷ However, the influence of H₂S is not limited to erectile functionality. Research led by Wang et al. highlighted its pivotal role in testicular function, especially spermatogenesis.⁸⁸ Conditions inducing stress, which resulted in decreased endogenous H₂S production and CBS expression in testes, led to hindered spermatogenesis and a disrupted blood-testicular barrier. Remarkably, introducing H₂S exogenously displayed therapeutic properties. H₂S provided protection against lipopolysaccharide-induced testicular abnormalities and rectified spermatogenic deficiencies. The primary mechanisms for these protective actions were attributed to anti-inflammatory and antioxidative properties of H₂S.⁸⁸⁻⁹³

At the molecular level, the transcription factors nuclear factor-κB (NF-κB) and nuclear factor erythroid 2 related factor 2 (Nrf2) play a decisive role in governing function of H₂S in reproduction.⁸⁸ Along with the mitogen-activated protein kinase (MAPK) signaling cascade, these transcription factors form the regulatory framework that underlies the protective influence of H₂S on male reproductive functions. Additionally, H₂S is essential for sperm motility and overall reproductive capability. Lower levels of H₂S in seminal plasma were observed in individuals with subfertility and infertility in comparison to fertile individuals.⁸⁸ This deficiency was not merely a chemical imbalance but had physiological manifestations as diminished CBS protein expression in sperm cells of asthenospermic and oligoasthenozoospermic subjects. Given the characteristic reduced sperm motility in these subjects, this highlights the vital role of H₂S in maintaining the fertilization capacity of sperm.⁸⁸ Supporting this notion was a study by Ikeda et al., which established a positive relationship between polysulfide concentrations in seminal fluid and sperm motility⁹⁴, suggesting the potential influence of polysulfide redox activity on

sperm function. Thus, hydrogen sulfide, an RSS, has been recognized as a key molecule in various facets of male reproductive health, ranging from erectile function, spermatogenesis, to sperm motility. Its interplay with different transcription factors and signaling cascades offers a deeper insight into its intricate role in male reproductive physiology. Continued exploration in this field promises to shed more light on its potential and diverse therapeutic applications.

Pathophysiological Significance of RSS in Reproductive Functions of Males

RSS, predominantly characterized by hydrogen sulfide (H_2S), have been identified as pivotal elements in male reproductive biology, drawing parallels to well-known reactive oxygen species (ROS) and RNS. Nonetheless, the precise equilibrium in the levels of these molecules, particularly H_2S , determines whether they play a beneficial or detrimental role in male reproductive health. Once viewed solely as a hazardous gas, H_2S was initially linked to harmful effects across multiple organ systems, eclipsing its possible advantageous impacts.⁹⁵ Current scientific interpretations, however, reveal that similar to ROS and RNS, the bifurcated outcomes of RSS (specifically H_2S) are contingent on concentration. Excessive H_2S concentrations, notably those beyond physiological thresholds, can be injurious to sperm functions.⁸⁸ For instance, interventions using sodium hydrosulfide ($NaHS$) have been found to markedly reduce sperm motility and exert cytotoxic effects on human sperm. Likewise, sodium sulfide (Na_2S), another H_2S donor, also inhibited sperm movement in both in vitro and in vivo studies.⁹⁶ At the core of this detrimental effect is the release of the gasotransmitter by certain H_2S donors in amounts exceeding physiological levels, which has negative repercussions for sperm functionality. This toxic action primarily stems from the obstruction of mitochondrial complex IV, leading to the halt of mitochondrial electron transport and the synthesis of sperm ATP.⁹⁷ As a result, the energy required for sperm motion becomes scarce, invariably compromising sperm motility and potentially leading to infertility.^{46, 98}

On a molecular level, in vitro studies using boar sperm revealed that H_2S interferes with multiple signaling cascades, subsequently compromising sperm movement.⁹⁶ Key molecular impacts include decreased ATP synthesis, activation of AMP-activated protein kinase (AMPK) and phosphatase and tensin homolog deleted on chromosome 10 (PTEN), and an increase in ROS concentrations. Elevated ROS levels might activate PTEN, which in turn might inhibit protein kinase B. The combined effects of ATP shortage and protein kinase B suppression activate AMPK, resulting in reduced sperm movement and, by extension, decreased sperm quality. Confirming these results, in vivo experiments using mouse sperm mirrored the in vitro findings.⁹⁶ On the other hand, a lack of H_2S in reproductive systems has also been associated with male infertility. A striking observation found that the CBS protein concentrations in the sperm of asthenospermic patients were notably lower than in normospermic counterparts, highlighting the crucial function of H_2S in maintaining sperm motility.⁸⁸ Additionally, an H_2S shortage in human plasma and corpus cavernosum samples was linked to a significant intensification of erectile dysfunction.⁹⁹ A decline in testosterone due to aging was concurrently linked to a drop in H_2S

production and concentrations in penile tissues, aligning with the symptomatic occurrences in erectile dysfunction.⁹⁹ Even with the knowledge acquired regarding the diverse roles of H_2S across both physiological and pathological domains, the holistic mechanism of action of H_2S remains elusive. As researchers continue to probe the nuances of H_2S in male reproductive physiology, it is crucial to recognize the delicate balance between its advantageous and disadvantageous effects, depending on its concentration and the specific scenario.^{100,101}

CONCLUSION

The complex interaction between RNS and RSS in male reproductive biology is a fascinating field of study. RNS and RSS play dual roles in both maintaining physiological processes and instigating pathological disturbances, highlighting their importance. Under normal conditions, they regulate key aspects of male fertility, such as sperm movement, capacitation, and the acrosome response. Their involvement in redox signaling serves as a critical regulator, ensuring cellular equilibrium and functionality. Conversely, an overabundance or imbalance can induce oxidative stress, which is linked to male infertility. Excessive levels of RNS and RSS can cause DNA lesions, lipid oxidation, and protein alterations, ultimately degrading sperm health and performance. Furthermore, new insights into the interaction between RNS and RSS shed light on the intertwined nature of redox pathways in the male reproductive system. Discrepancies in this interaction could heighten or predispose the adverse effects on male reproductive capabilities, underscoring the importance of interventions aimed at these reactive species. Future research will greatly benefit from a detailed examination of the molecular pathways influenced by these reactive species and potential measures to regulate their concentrations. As our comprehension of RNS and RSS expands, recognizing their dual functions becomes vital. This knowledge allows us to leverage their physiological advantages while counteracting their pathological threats, presenting potential solutions to male reproductive issues.

Conflict of Interest

The authors declare that none of them has any conflict of interest.

Funding: None

REFERENCES

1. A. Agarwal, P. Sengupta. Oxidative stress and its association with male infertility. *Male infertility: contemporary clinical approaches, andrology, ART and antioxidants* **2020**, 57-68.
2. S. Dutta, R. Henkel, P. Sengupta, A. Agarwal. Physiological role of ROS in sperm function. *Male infertility: Contemporary clinical approaches, Andrology, ART and antioxidants* **2020**, 337-345.
3. A. Agarwal, K. Leisegang, P. Sengupta, *Oxidative stress in pathologies of male reproductive disorders*, in *Pathology*. 2020, Elsevier. p. 15-27.
4. M.C. Gruhke, A.J. Slusarenko. The biology of reactive sulfur species (RSS). *Plant Physiol Biochem* **2012**, 59, 98-107.
5. S. Dutta, P. Sengupta, S. Das, P. Slama, S. Roychoudhury. Reactive Nitrogen Species and Male Reproduction: Physiological and Pathological Aspects. *Int J Mol Sci* **2022**, 23(18), 10574.
6. M.J. Tomlinson, S.J. East, C.L. Barratt, A.E. Bolton, I.D. Cooke. Preliminary communication: possible role of reactive nitrogen intermediates in leucocyte-mediated sperm dysfunction. *Am J Reprod Immunol* **1992**, 27(1-2), 89-92.

7. S. Dutta, P. Sengupta. Role of nitric oxide in male and female reproduction. *Malays J Med Sci* **2021**.
8. S. Dutta, P. Sengupta, P. Slama, S. Roychoudhury. Oxidative Stress, Testicular Inflammatory Pathways, and Male Reproduction. *Int J Mol Sci* **2021**, 22(18).
9. V. Otasevic, A. Stancic, A. Korac, A. Jankovic, B. Korac. Reactive oxygen, nitrogen, and sulfur species in human male fertility. A crossroad of cellular signaling and pathology. *Biofactors* **2020**, 46(2), 206-219.
10. W. Alderton, C. Cooper, R. Knowles. Nitric oxide synthases: structure, function and inhibition. *Biochem J* **2001**, 357, 593-615.
11. R. Govers, S. Oess. To NO or not to NO, where, is the question. *Histol Histopathol* **2004**.
12. V. Otasevic, A. Korac, B. Buzadzic, A. Stancic, A. Jankovic, B. Korac. Nitric oxide and thermogenesis-challenge in molecular cell physiology. *Front Biosci* **2011**, 3(3), 1180-1195.
13. A. Martínez-Ruiz, S. Cadenas, S. Lamas. Nitric oxide signaling: classical, less classical, and nonclassical mechanisms. *Free Rad Biol Med* **2011**, 51(1), 17-29.
14. J.J. Mיעyal, M.M. Gallogly, S. Qanungo, E.A. Sabens, M.D. Shelton. Molecular mechanisms and clinical implications of reversible protein S-glutathionylation. *Antiox Redox Signal* **2008**, 10(11), 1941-1988.
15. J.M. Souza, G. Peluffo, R. Radi. Protein tyrosine nitration—functional alteration or just a biomarker? *Free Rad Biol Med* **2008**, 45(4), 357-366.
16. J. Stamler. S-nitrosothiols and the bioregulatory actions of nitrogen oxides through reactions with thiol groups. *The role of nitric oxide in physiology and pathophysiology* **1995**, 19-36.
17. R. Radi. Oxygen radicals, nitric oxide, and peroxynitrite: Redox pathways in molecular medicine. *Proc Nat Acad Sci* **2018**, 115(23), 5839-5848.
18. C. Lindermayr, G. Saalbach, J.r. Durner. Proteomic identification of S-nitrosylated proteins in Arabidopsis. *Plant Physiol* **2005**, 137(3), 921-930.
19. H. Oberkofler, G. Dallinger, Y. Liu, E. Hell, F. Krempler, W. Patsch. Uncoupling protein gene: quantification of expression levels in adipose tissues of obese and non-obese humans. *J Lipid Res* **1997**, 38(10), 2125-2133.
20. A. Jankovic, A. Korac, B. Buzadzic, A. Stancic, V. Otasevic, P. Ferdinandy, et al. Targeting the NO/superoxide ratio in adipose tissue: relevance to obesity and diabetes management. *Brit J Pharmacol* **2017**, 174(12), 1570-1590.
21. M.B. Herrero, E. de Lamirande, C. Gagnon. Tyrosine nitration in human spermatozoa: a physiological function of peroxynitrite, the reaction product of nitric oxide and superoxide. *Mol Hum Reprod* **2001**, 7(10), 913-921.
22. E. de Lamirande, G. Lamothe, M. Villemure. Control of superoxide and nitric oxide formation during human sperm capacitation. *Free Radic Biol Med* **2009**, 46(10), 1420-1427.
23. A. Zini, M.K. O'Bryan, M.S. Magid, P.N. Schlegel. Immunohistochemical localization of endothelial nitric oxide synthase in human testis, epididymis, and vas deferens suggests a possible role for nitric oxide in spermatogenesis, sperm maturation, and programmed cell death. *Biol Reprod* **1996**, 55(5), 935-941.
24. N.P. Lee, C. Yan Cheng. Regulation of Sertoli cell tight junction dynamics in the rat testis via the nitric oxide synthase/soluble guanylate cyclase/3', 5'-cyclic guanosine monophosphate/protein kinase G signaling pathway: an in vitro study. *Endocrinology* **2003**, 144(7), 3114-3129.
25. M.K. O'Bryan, A. Zini, C.Y. Cheng, P.N. Schlegel. Human sperm endothelial nitric oxide synthase expression: correlation with sperm motility. *Fertil Steril* **1998**, 70(6), 1143-1147.
26. K.-E. Andersson. Pharmacology of erectile function and dysfunction. *Urol Clin North Am* **2001**, 28(2), 233-248.
27. C. Roessner, U. Paasch, H.J. Glander, S. Grunewald. Activity of nitric oxide synthase in mature and immature human spermatozoa. *Andrologia* **2010**, 42(2), 132-137.
28. M.E. Sullivan, C.S. Thompson, M.R. Dashwood, M.A. Khan, J.Y. Jeremy, R.J. Morgan, et al. Nitric oxide and penile erection: is erectile dysfunction another manifestation of vascular disease? *Cardiovas Res* **1999**, 43(3), 658-665.
29. G. Yetik-Anacak, R. Sorrentino, A. Linder, N. Murat. Gas what: NO is not the only answer to sexual function. *Brit J Pharmacol* **2015**, 172(6), 1434-1454.
30. A.L. Burnett, A.G. Chang, J.K. Crone, P.L. Huang, S.F. SEZEN. Noncholinergic penile erection in mice lacking the gene for endothelial nitric oxide synthase. *J Androl* **2002**, 23(1), 92-97.
31. D.E. Cashen, D.E. MacIntyre, W.J. Martin. Effects of sildenafil on erectile activity in mice lacking neuronal or endothelial nitric oxide synthase. *Brit J Pharmacol* **2002**, 136(5), 693.
32. G.F. Lasker, C.J. Matt, J. Badejo, Adeleke M, D.B. Casey, J.S. Dhaliwal, S.N. Murthy, et al. Intracavernosal administration of sodium nitrite as an erectile pharmacotherapy. *Canad J Physiol Pharmacol* **2010**, 88(7), 770-776.
33. S. Kothari, A. Thompson, A. Agarwal, S.S. du Plessis. Free radicals: their beneficial and detrimental effects on sperm function. *Indian J Exp Biol* **2010**, 48(5), 425-435.
34. E. Salvolini, E. Buldreghini, G. Lucarini, A. Vignini, R. Di Primio, G. Balercia. Nitric oxide synthase and tyrosine nitration in idiopathic asthenozoospermia: an immunohistochemical study. *Fertil Steril* **2012**, 97(3), 554-560.
35. A. Cassina, P. Silveira, L. Cantu, J.M. Montes, R. Radi, R. Sapiro. Defective Human Sperm Cells Are Associated with Mitochondrial Dysfunction and Oxidant Production. *Biol Reprod* **2015**, 93(5), 119.
36. P.C. Rodriguez, M.T. Beconi. Peroxynitrite participates in mechanisms involved in capacitation of cryopreserved cattle. *Anim Reprod Sci* **2009**, 110(1-2), 96-107.
37. T. Nobunaga, Y. Tokugawa, K. Hashimoto, Y. Kubota, K. Sawai, T. Kimura, et al. Elevated nitric oxide concentration in the seminal plasma of infertile males: nitric oxide inhibits sperm motility. *Am J Reprod Immunol* **1996**, 36(4), 193-197.
38. M. Rosselli, R.K. Dubey, B. Imthurn, E. Macas, P.J. Keller. Effects of nitric oxide on human spermatozoa: evidence that nitric oxide decreases sperm motility and induces sperm toxicity. *Hum Reprod* **1995**, 10(7), 1786-1790.
39. M. Rosselli, R.K. Dubey, B. Imthurn, E. Macas, P.J. Keller. Andrology: Effects of nitric oxide on human spermatozoa: Evidence that nitric oxide decreases sperm motility and induces sperm toxicity. *Hum Reprod* **1995**, 10(7), 1786-1790.
40. W.J. Hellstrom, M. Bell, R. Wang, S.C. Sikka. Effect of sodium nitroprusside on sperm motility, viability, and lipid peroxidation. *Fertil Steril* **1994**, 61(6), 1117-1122.
41. E.T. Donnelly, S.E. Lewis, W. Thompson, U. Chakravarthy. Sperm nitric oxide and motility: the effects of nitric oxide synthase stimulation and inhibition. *Mol Hum Reprod* **1997**, 3(9), 755-762.
42. R.R. Yeoman, W.D. Jones, B.M. Rizk. Evidence for nitric oxide regulation of hamster sperm hyperactivation. *J Androl* **1998**, 19(1), 58-64.
43. S. Lewis, E. Donnelly, E. Sterling, M. Kennedy, W. Thompson, U. Chakravarthy. Nitric oxide synthase and nitrite production in human spermatozoa: evidence that endogenous nitric oxide is beneficial to sperm motility. *Mol Hum Reprod* **1996**, 2, 873-878.
44. J.P. Bolaños, M. Delgado-Esteban, A. Herrero-Mendez, S. Fernandez-Fernandez, A. Almeida. Regulation of glycolysis and pentose-phosphate pathway by nitric oxide: Impact on neuronal survival. *Biochim Biophys Acta* **2008**, 1777(7-8), 789-793.
45. E. Miraglia, F. De Angelis, E. Gazzano, H. Hassanpour, A. Bertagna, E. Aldieri, et al. Nitric oxide stimulates human sperm motility via activation of the cyclic GMP/protein kinase G signaling pathway. *Reproduction* **2011**, 141(1), 47-54.
46. V. Otasevic, A. Korac, M. Vucetic, B. Macanovic, E. Garalejic, I. Ivanovic-Burmazovic, et al. Is manganese (II) pentaazamacrocyclic superoxide dismutase mimic beneficial for human sperm mitochondria function and motility? *Antioxid Redox Signal* **2013**, 18(2), 170-178.
47. P. Sengupta. Reviewing reports of semen volume and male aging of last 33 years: From 1980 through 2013. *Asian Pac J Reprod* **2015**, 4(3), 242-246.
48. A. Revelli, C. Costamagna, F. Moffa, E. Aldieri, S. Ochetti, A. Bosia, et al. Signaling pathway of nitric oxide-induced acrosome reaction in human spermatozoa. *Biol Reprod* **2001**, 64(6), 1708-1712.

49. A. Zini, E. de Lamirande, C. Gagnon. Low levels of nitric oxide promote human sperm capacitation in vitro. *J Androl* **1995**, 16(5), 424-431.
50. M.B. Herrero, E. de Lamirande, C. Gagnon. Nitric oxide regulates human sperm capacitation and protein-tyrosine phosphorylation in vitro. *Biol Reprod* **1999**, 61(3), 575-581.
51. C. O'Flaherty, E. de Lamirande, C. Gagnon. Positive role of reactive oxygen species in mammalian sperm capacitation: triggering and modulation of phosphorylation events. *Free Rad Biol Med* **2006**, 41(4), 528-540.
52. E. de Lamirande, C. O'Flaherty. Sperm activation: role of reactive oxygen species and kinases. *Biochim Biophys Acta* **2008**, 1784(1), 106-115.
53. C. Sezer, I. Koksak, M. Usta, K. Gulkesen, T. Erdogru, A. Ciftcioglu, et al. Relationship between mast cell and iNOS expression in testicular tissue associated with infertility. *Arch Androl* **2005**, 51(2), 149-158.
54. E. Miraglia, M.L. Rullo, A. Borgia, M. Massobrio, A. Revelli, D. Ghigo. Stimulation of the nitric oxide/cyclic guanosine monophosphate signaling pathway elicits human sperm chemotaxis in vitro. *Fertil Steril* **2007**, 87(5), 1059-1063.
55. N. Gonzalez-Cadavid, J. Rajfer. The pleiotropic effects of inducible nitric oxide synthase (iNOS) on the physiology and pathology of penile erection. *Curr Pharm Des* **2005**, 11(31), 4041-4046.
56. G. Balercia, S. Moretti, A. Vignini, M. Magagnini, F. Mantero, M. Boscaro, et al. Role of nitric oxide concentrations on human sperm motility. *J Androl* **2004**, 25(2), 245-249.
57. A. Semenova, I. Tomilova, K. Panikratov, E. Kadykova, A. Basharin. The role of nitric oxide in fertility disorders in men. *Urologia* **2005**(6), 31-36.
58. E. de Lamirande, C. Gagnon. Capacitation-associated production of superoxide anion by human spermatozoa. *Free Radic Biol Med* **1995**, 18(3), 487-495.
59. U. Förstermann, E.I. Closs, J.S. Pollock, M. Nakane, P. Schwarz, I. Gath, et al. Nitric oxide synthase isozymes. Characterization, purification, molecular cloning, and functions. *Hypertension* **1994**, 23(6_pt_2), 1121-1131.
60. K. Sengoku, K. Tamate, T. Yoshida, Y. Takaoka, T. Miyamoto, M. Ishikawa. Effects of low concentrations of nitric oxide on the zona pellucida binding ability of human spermatozoa. *Fertil Steril* **1998**, 69(3), 522-527.
61. F. Francavilla, R. Santucci, B. Macerola, G. Ruvolo, R. Romano. Nitric oxide synthase inhibition in human sperm affects sperm-oocyte fusion but not zona pellucida binding. *Biol Reprod* **2000**, 63(2), 425-429.
62. A. Revelli, G. Soldati, C. Costamagna, O. Pellerey, E. Aldieri, M. Massobrio, et al. Follicular fluid proteins stimulate nitric oxide (NO) synthesis in human sperm: a possible role for NO in acrosomal reaction. *J Cell Physiol* **1999**, 178(1), 85-92.
63. M.B. Herrero, E. de Lamirande, C. Gagnon. Nitric oxide is a signaling molecule in spermatozoa. *Curr Pharm Des* **2003**, 9(5), 419-425.
64. N.V. Hud, M.J. Allen, K.H. Downing, J. Lee, R. Balhorn. Identification of the elemental packing unit of DNA in mammalian sperm cells by atomic force microscopy. *Biochem Biophys Res Commun* **1993**, 193(3), 1347-1354.
65. M.M. Cortese-Krott, A. Koning, G.G. Kuhnle, P. Nagy, C.L. Bianco, A. Pasch, et al. The reactive species interactome: evolutionary emergence, biological significance, and opportunities for redox metabolomics and personalized medicine. *Antioxid Redox Signal* **2017**, 27(10), 684-712.
66. J.B. Weinberg, E. Doty, J. Bonaventura, A. Haney. Nitric oxide inhibition of human sperm motility. *Fertil Steril* **1995**, 64(2), 408-413.
67. S. Dutta, P. Sengupta, S. Muhamad. Male reproductive hormones and semen quality. *Asian Pac J Reprod* **2019**, 8(5).
68. R.K. Sharma, A. Agarwal. Role of reactive oxygen species in male infertility. *Urology* **1996**, 48(6), 835-850.
69. T.P. Wu, B.M. Huang, H.C. Tsai, M.C. Lui, M.Y. Liu. Effects of nitric oxide on human spermatozoa activity, fertilization and mouse embryonic development. *Arch Androl* **2004**, 50(3), 173-179.
70. T. Kostic, S. Andric, R. Kovacevic, D. Maric. Is nitric oxide involved in stress impaired testicular steroidogenesis? *Zbornik Matice Srpske Za Prirodne Nauke* **1998**.
71. R. Middendorff, D. Muller, S. Wichers, A. Holstein, M. Davidoff. Evidence for production and functional activity of nitric oxide in seminiferous tubules and blood vessels of the human testis. *J Clin Endocrinol Metab* **1997**, 82(12), 4154-4161.
72. S. Miyamoto, H. Teramoto, O.A. Coso, J.S. Gutkind, P.D. Burbelo, S.K. Akiyama, et al. Integrin function: molecular hierarchies of cytoskeletal and signaling molecules. *J Cell Biol* **1995**, 131(3), 791-805.
73. G. Santoro, C. Romeo, P. Impellizzeri, R. Ientile, G. Cutroneo, F. Trimarchi, et al. Nitric oxide synthase patterns in normal and varicocele testis in adolescents. *BJU Int* **2001**, 88(9), 967-973.
74. J. Yang, T. Wang, J. Yang, K. Rao, Y. Zhan, R.B. Chen, et al. S-allyl cysteine restores erectile function through inhibition of reactive oxygen species generation in diabetic rats. *Andrology* **2013**, 1(3), 487-494.
75. Y. Aksoy, I. Özbey, H. Aksoy, Ö. Polat, F. Akca. Seminal plasma nitric oxide concentration in oligo-and/or asthenozoospermic subjects with/without varicocele. *Arch Androl* **2002**, 48(3), 181-185.
76. I. Huang, J. Jones, O. Khorram. Human seminal plasma nitric oxide: correlation with sperm morphology and testosterone. *Med Sci Monit* **2006**, 12(3), Cr103-106.
77. H.P. Monteiro. Signal transduction by protein tyrosine nitration: competition or cooperation with tyrosine phosphorylation-dependent signaling events? *Free Radic Biol Med* **2002**, 33(6), 765-773.
78. T. Ramya, M.M. Misro, D. Sinha, D. Nandan, S. Mithal. Altered levels of seminal nitric oxide, nitric oxide synthase, and enzymatic antioxidants and their association with sperm function in infertile subjects. *Fertil Steril* **2011**, 95(1), 135-140.
79. R. Wang. Physiological implications of hydrogen sulfide: a whiff exploration that blossomed. *Physiol Rev* **2012**, 92(2), 791-896.
80. O. Kabil, N. Motl, R. Banerjee. H₂S and its role in redox signaling. *Biochim Biophys Acta* **2014**, 1844(8), 1355-1366.
81. P. Nagy, Z. Pálkás, A. Nagy, B. Budai, I. Tóth, A. Vasas. Chemical aspects of hydrogen sulfide measurements in physiological samples. *Biochim Biophys Acta* **2014**, 1840(2), 876-891.
82. R. Reiffenstein, W.C. Hulbert, S.H. Roth. Toxicology of hydrogen sulfide. *Ann Rev Pharmacol Toxicol* **1992**, 32(1), 109-134.
83. Y. Sugiura, M. Kashiba, K. Maruyama, K. Hoshikawa, R. Sasaki, K. Saito, et al. Cadmium exposure alters metabolomics of sulfur-containing amino acids in rat testes. *Antioxid Redox Signal* **2005**, 7(5-6), 781-787.
84. P. Patel, M. Vatis, J. Heptinstall, R. Wang, R.J. Carson. The endogenous production of hydrogen sulphide in intrauterine tissues. *Reprod Biol Endocrinol* **2009**, 7(1), 1-9.
85. X.-Y. Zhu, H. Gu, X. Ni. Hydrogen sulfide in the endocrine and reproductive systems. *Expert Rev Clin Pharmacol* **2011**, 4(1), 75-82.
86. B. Srilatha, P.G. Adaikan, P.K. Moore. Possible role for the novel gasotransmitter hydrogen sulphide in erectile dysfunction—A pilot study. *Eur J Pharmacol* **2006**, 535(1-3), 280-282.
87. R. d'Emmanuele di Villa Bianca, R. Sorrentino, P. Maffia, V. Mirone, C. Imbimbo, F. Fusco, et al. Hydrogen sulfide as a mediator of human corpus cavernosum smooth-muscle relaxation. *Proc Nat Acad Sci* **2009**, 106(11), 4513-4518.
88. J. Wang, W. Wang, S. Li, Y. Han, P. Zhang, G. Meng, et al. Hydrogen sulfide as a potential target in preventing spermatogenic failure and testicular dysfunction. *Antioxid Redox Signal* **2018**, 28(16), 1447-1462.
89. T. Irez, S. Bicer, E. Sahin, S. Dutta, P. Sengupta. Cytokines and adipokines in the regulation of spermatogenesis and semen quality. *Chem Biol Lett* **2020**, 7(2), 131-139.
90. K. Bhattacharya, P. Sengupta, S. Dutta, I.R. Karkada. Obesity, systemic inflammation and male infertility. *Chem Biol Lett* **2020**, 7(2), 92-98.
91. S. Dutta, A. Biswas, P. Sengupta. Obesity, endocrine disruption and male infertility. *Asian Pac J Reprod* **2019**, 8(5), 195.
92. P. Sengupta, S. Dutta, A.T. Alahmar, U.J.A. D'souza. Reproductive tract infection, inflammation and male infertility. *Chema Biol Lett* **2020**, 7(2), 75-84.
93. S. Dutta, P. Sengupta, B.S. Chhikara. Reproductive inflammatory mediators and male infertility. *Chem Biol Lett* **2020**, 7(2), 73-74.
94. M. Ikeda, Y. Ishima, V.T. Chuang, M. Sakai, H. Osafune, H. Ando, et al. Distribution of polysulfide in human biological fluids and their association with amylase and sperm activities. *Molecules* **2019**, 24(9), 1689.

95. H. Kimura. Production and physiological effects of hydrogen sulfide. *Antioxid Redox Signal* **2014**, 20(5), 783-793.
96. Y. Zhao, W.D. Zhang, X.Q. Liu, P.F. Zhang, Y.-N. Hao, L. Li, et al. Hydrogen sulfide and/or ammonia reduces spermatozoa motility through AMPK/AKT related pathways. *Sci Reports* **2016**, 6(1), 37884.
97. C. Szabo, C. Ransy, K. Módis, M. Andriamihaja, B. Murgheș, C. Coletta, et al. Regulation of mitochondrial bioenergetic function by hydrogen sulfide. Part I. Biochemical and physiological mechanisms. *Brit J Pharmacol* **2014**, 171(8), 2099-2122.
98. F. Gallon, C. Marchetti, N. Jouy, P. Marchetti. The functionality of mitochondria differentiates human spermatozoa with high and low fertilizing capability. *Fertil Steril* **2006**, 86(5), 1526-1530.
99. B. Srilatha, P. Muthulakshmi, P.G. Adaikan, P.K. Moore. Endogenous hydrogen sulfide insufficiency as a predictor of sexual dysfunction in aging rats. *Aging Male* **2012**, 15(3), 153-158.
100. P. Lakra, I.N. Gahlawat. Regular food chemicals as antioxidant towards prevention of diseases – An insight review. *J. Mol. Chem.* **2022**, 2 (2), 441.
101. C. Pal. Molecular mechanism facets of Oxidative stress mediated pathogenesis. *J. Mol. Chem.* **2023**, 3 (2), 587.