

Development and optimization of *Andrographis paniculata* extract-loaded phytosomes using Box-Behnken design approach

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ABSTRACT

In current research, *Andrographis paniculata* extracts loaded phytosomes were successfully synthesized by thin-film hydration technique using soy lecithin and cholesterol and optimized by Box-Behnken design. The quadratic equation for % entrapment efficiency was found $Y_1 = 69.22 + 15.23 X_1 - 0.7775 X_2 - 2.65 X_3 - 1.025 X_1 X_2 + 2.41 X_1 X_3 - 1.56 X_2 X_3 + 6.95 X_1^2 - 7.21 X_2^2 - 5.52 X_3^2$ and for percentage yield was found $Y_2 = 70.43 + 13.86 X_1 - 0.2025 X_2 - 0.6962 X_3 - 0.2425 X_1 X_2 + 1.86 X_1 X_3 - 1.43 X_2 X_3 + 6.47 X_1^2 - 6.75 X_2^2 - 3.83 X_3^2$. These equations demonstrated that lipid: drug (X_1) have significant effect on entrapment efficiency (Y_1) and yield (Y_2). The values of independent variables for optimized phytosomes were lipid: extract ($X_1 = 1: 1$ w/w), temperature ($X_2 = 43^\circ\text{C}$) and time ($X_3 = 2.15$ hours) which have D-value of 0.929. The % bias between actual and predicted values of Y_1 and Y_2 was 1.25% and 2.36%, respectively which concluded authenticity of design model.

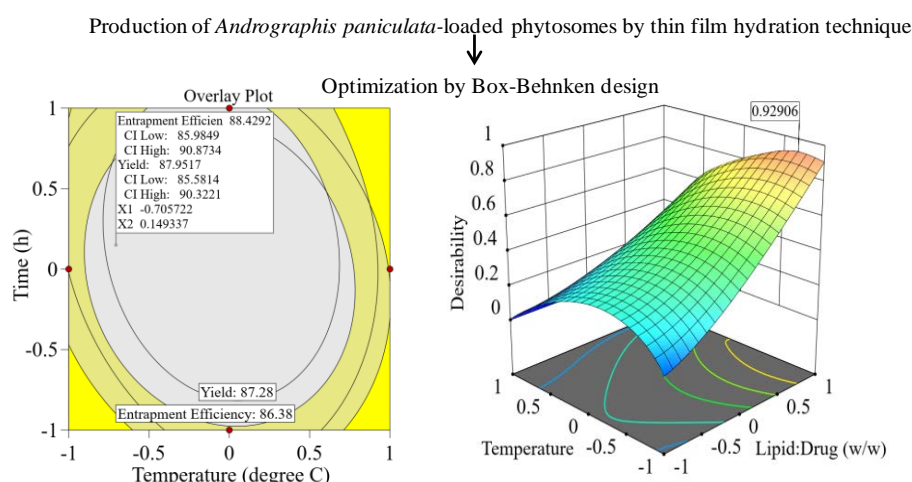
Keywords: *Andrographis paniculata*, Box-Behnken Design, Phytosomes, Thin-Film Hydration, Entrapment Efficiency

INTRODUCTION

Liver damage and malfunction starts with small incidence of damage to hepatocytes by different factors including viral, bacterial or other infections, alcoholic, drug induced or fatty damage leading to inflammatory changes causing hepatitis, which may start as acute change to chronic hepatitis, gradually accumulating fibrous tissue around necrotized areas and advancing to fibrosis and steatosis which may gradually become irreversible as cirrhosis.¹ All liver changes are well manifested by changes in hepatic enzyme levels, serum bilirubin and biliverdin and antioxidant capacity. Liver has a

capacity to regenerate until the condition changes to fibrosis and cirrhosis, when bile duct and hepatic duct structures are damaged creating changes in serum biliary enzyme content which gets evident as jaundice. Various synthetic medicinal agents have been tried, but a comprehensive herbal have patient compliant remains the need of the hour.² Phytosome technology can be explored for incorporation of standardized plant extracts or water-insoluble phytoconstituents into phospholipids to produce lipid compatible molecular complexes which improves their absorption and bioavailability.³ Numerous plant extracts and phytoconstituents have been reported for hepatoprotective activity.⁴⁻⁶ This has been observed that phytoconstituents act in synergism, and so it is relevant that a combination formulation can be formulated and screened for getting the best therapeutic alternative.^{7,8}

In this research, *Andrographis paniculata*-loaded phytosomes were synthesized through thin film hydration technique using soy lecithin and cholesterol and formulation optimization was executed by Box-Behnken design. The objective of this research was to



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examine influence of factors on percentage entrapment efficiency and percentage yield of extract-loaded phytosomes. The statistical and diagnostic analysis of dependent variables was done to inspect model fit and to confirm reliability of model, respectively. The optimized extract-loaded phytosomes was explored by Design-Expert software and model validation was executed by check point analysis.

MATERIALS AND METHODS

Andrographis paniculata extract was procured from NJP Healthcare Pvt. Ltd. Gujarat. Soy lecithin and cholesterol were procured from Loba Chemicals Private Limited, Mumbai, India. All other ingredients employed were of analytical grade.

Experimental design

Three-factor Box-Behnken design was used as experimental design layout for manufacturing of phytosomes. Independent and dependent parameters investigated during this research have been depicted in Table 1.⁹⁻¹³

Table 1. Independent and dependent variables studied during synthesis of extract-loaded phytosomes

Independent variables	Levels of variables		
	-1	0	+1
Lipid: Extract (w/w) (X1)	0.5:1	1:1	1.5:1
Temperature (°C) (X2)	40	50	60
Time (hrs) (X3)	1	2	3
Dependent variables	Constraint		
Entrapment Efficiency (% w/w) (Y1)	Maximize		
Yield (% w/w) (Y2)	Maximize		

Production of extract-loaded phytosomes

The phytosomes were synthesized according to previously described procedure.¹⁴⁻¹⁷ Extract and soy lecithin in different ratios were transferred into round bottom flask and cholesterol was mixed to above mixture which was subsequently refluxed with 20 ml dichloromethane at specific temperatures for various time intervals (Table 2). The mixture was evaporated off under vacuum to produce thin film which was hydrated with 50 mL phosphate buffer having pH 7.4. The dispersion was homogenized using probe sonicator (Ultrasonic probe sonicator, PCI analytics) at 230 voltage and 50 Hz AC current at 20 seconds, filtered using membrane filtration using 0.45 µm filter and transferred into glass vial for storage at room temperature.

Evaluation of response variables of extract-loaded phytosomes

The percentage entrapment efficiency of phytosomes was evaluated in terms of andrographolides contents within phytosomes. The phytosomes were subjected to ultracentrifuge and subsequently supernatants were kept aside for twenty minutes and analysed using UV-visible spectrophotometer at 494 nm. The percentage entrapment efficiency was determined using equation 1.^{18,19}

$$\% \text{ Entrapment efficiency} = [(TAC-DAC)/TAC]*100 \quad \text{Eq. 1}$$

Table 2. Box-Behnken design layout with experimental values of response variables of extract-loaded phytosomes (Batch 1-15)

Batch	Independent variables			Dependent variables	
	X1	X2	X3	Y1	Y2
1	-1	-1	0	53.22	56.65
2	1	-1	0	86.38	85.23
3	-1	1	0	51.75	55.56
4	1	1	0	84.5	83.17
5	-1	0	-1	61.59	62.57
6	1	0	-1	84.73	86.21
7	-1	0	1	51.75	56.22
8	1	0	1	84.51	87.28
9	0	-1	-1	58.43	58.11
10	0	1	-1	60.11	61.73
11	0	-1	1	55.98	60.82
12	0	1	1	51.43	58.73
13	0	0	0	69.1	72.16
14	0	0	0	70.2	68.35
15	0	0	0	68.35	70.78

X1: Lipid: Drug (w/w); X2: Temperature (°C); X3: Time (hrs); Y1: EE (% w/w); Y2: Yield (% w/w)

Where, TAC and DAC are theoretical and detected andrographolides content, respectively. Percent yield (Y2) was determined as percentage weight fraction of phytosomes with initial total weight of extracts, soy lecithin and cholesterol (Eq. 2).²⁰⁻²²

$$\% \text{ Yield} = [W1/W2]*100 \quad \text{Eq. 2}$$

Where, 'W1' is final weight of phytosomes collected and 'W2' is initial weight of extract, soy lecithin and cholesterol.

Analysis of response variables by Design-Expert

The selection of appropriate model for response variables analysis was performed on the basis of sequential *p*-values and lack-of-fit *p*-values of linear, 2-FI (2-factors-interaction), quadratic and cubic model.²³⁻²⁵ Statistical analysis was executed by analysis of variance was executed to determine *p*-value for independent parameters. The diagnostic analysis was executed to check for outliers in design model. The contour and response surface plots were obtained which demonstrated graphical view of independent versus response variables.²⁶⁻²⁹

Optimization and validation of extract-loaded phytosomes

Optimal values of parameters for production of optimized batch of phytosome with highest overall desirability function (D-value) were obtained through analysis.^{30,31} The new check point batch of phytosome was synthesized to validate optimization strategy by estimating percentage bias using Eq. (3).

$$\% \text{ Bias} = [PV-EV/PV]*100 \quad \text{Eq. 3}$$

Where, PY is predicted value and EV is experimental value.

RESULTS AND DISCUSSION

Analysis of entrapment efficiency (% w/w) (Y1)

The quadratic model was selected for entrapment efficiency (Y1) since this was found that quadratic model has maximum *R*² value of 0.9942, insignificant lack of fit value of 0.1745 and sequential *p*-value of 0.0002 (Table 3). Figure 1 demonstrated

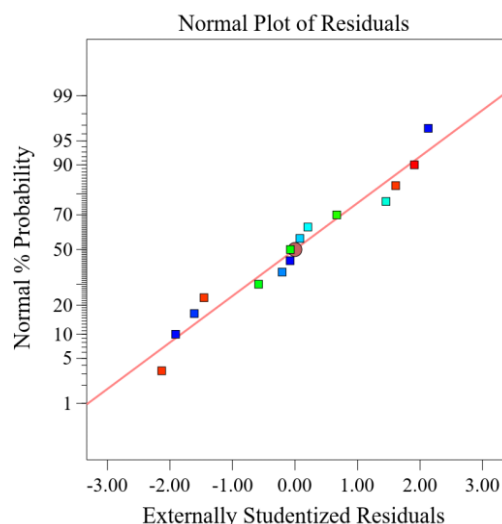


Figure 1. Normal probability plot of residuals for percentage entrapment efficiency

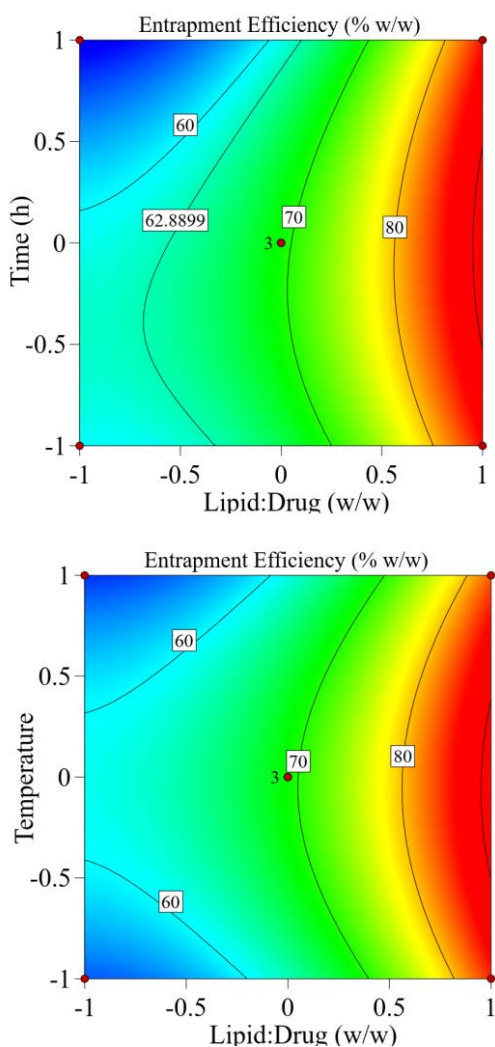


Figure 2. Illustration of effect of independent variables over percentage entrapment efficiency using two-dimensional contour plots

normal probability plot of residuals for percentage entrapment efficiency which revealed normality of data. Quadratic equation for percentage entrapment efficiency (Y1) developed by multiple regression analysis has been depicted below:

$$\% \text{ Entrapment efficiency (Y1)} = 69.22 + 15.23 X_1 - 0.7775 X_2 - 2.65 X_3 - 1.025 X_1 X_2 + 2.41 X_1 X_3 - 1.56 X_2 X_3 + 6.95 X_1^2 - 7.21 X_2^2 - 5.52 X_3^2$$

Eq. (4)

The negative value of coefficient for X2 and X3 revealed their antagonistic action while the positive value of coefficient for X1 demonstrated their synergistic influence on Y1 of phytosomes.³²⁻³⁵ The *p*-value for X1, X3, X1², X2² and X3² was less than 0.05 (Table 4) which illustrated that lipid: extract (X1) and time (X3) significantly affected entrapment efficiency (Y1) as indicated through response surface graphs (Figure 2 and 3).^{19-24,36} The increase in percentage entrapment efficiency with increase in lipid: extract from 0.25:1 to 1:1 might be due to increase in amount of lipid which provide more space to accommodate extract.^{11,37,38}

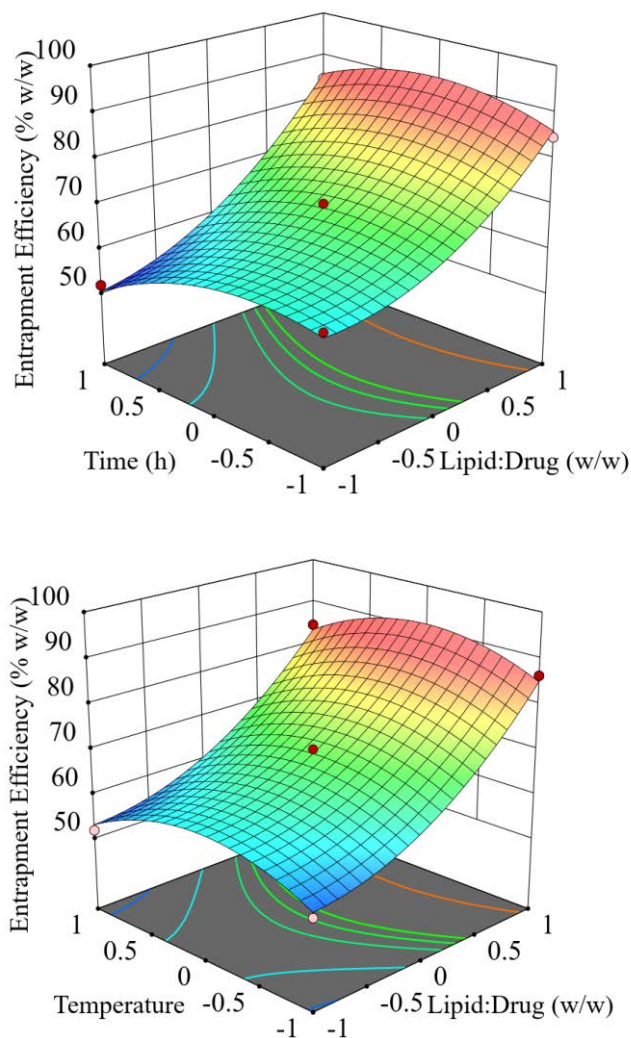


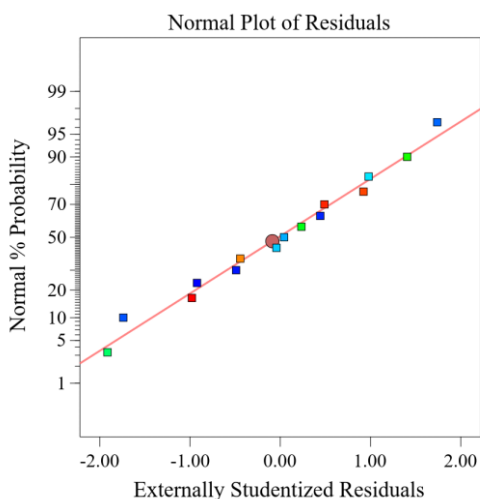
Figure 3. Illustration of effect of independent variables over percentage entrapment efficiency using three-dimensional response surface plots

Table 1. Fit summary and lack of fit statistics of Y1 and Y2

Source	Parameter	R ²	Lack of Fit <i>p</i> -value	Sequential <i>p</i> -value
Linear	Y1	0.7734	0.0138	0.0007
	Y2	0.7780	0.0739	0.0006
2FI	Y1	0.7866	0.0098	0.9164
	Y2	0.7892	0.0525	0.9323
Quadratic	Y1	0.9942	0.1745	0.0002
	Y2	0.9932	0.6965	0.0004
Cubic	Y1	0.9993	-	0.1745
	Y2	0.9962	-	0.6965

Table 2. Analysis of variance of percentage entrapment efficiency

Source	Sum of Squares	Df	Mean Square	F-value	p-value
Model	2462.68	9	273.63	94.82	< 0.0001
X ₁	1854.71	1	1854.71	642.72	< 0.0001
X ₂	4.84	1	4.84	1.68	0.2520
X ₃	56.13	1	56.13	19.45	0.0070
X ₁ X ₂	0.0420	1	0.0420	0.0146	0.9086
X ₁ X ₃	23.14	1	23.14	8.02	0.0366
X ₂ X ₃	9.70	1	9.70	3.36	0.1262
X ₁ ²	178.43	1	178.43	61.83	0.0005
X ₂ ²	191.72	1	191.72	66.44	0.0005
X ₃ ²	112.64	1	112.64	39.03	0.0015
Lack of fit	12.70	3	4.23	4.89	0.1745

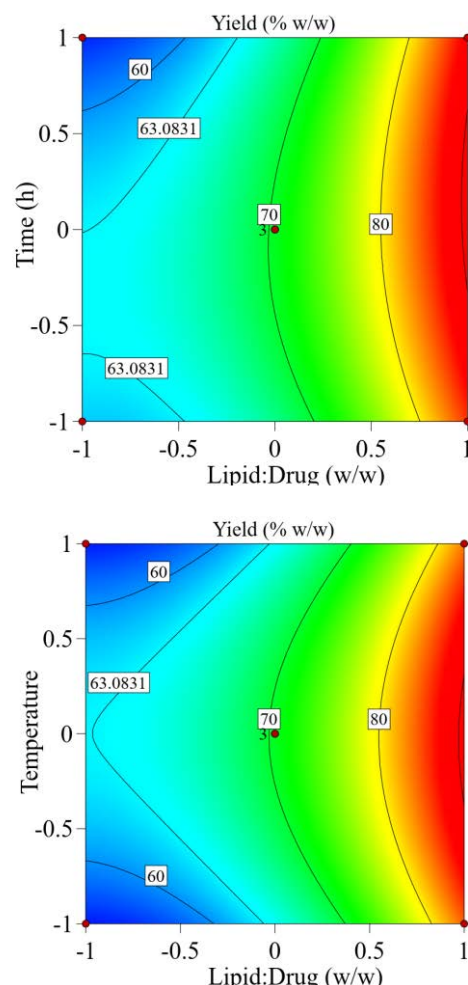
**Figure 4.** Normal probability plot of residuals for percentage entrapment efficiency

Analysis of yield (% w/w) (Y2)

The quadratic model was chosen for % yield (Y2) due to highest R² of 0.9932, insignificant ($p > 0.05$) lack of fit of 0.6965 and sequential p -value of 0.0004 (Table 3). Figure 4 revealed normal probability plot of residuals for percentage yield which revealed normality of percentage yield values. Quadratic equation for percentage yield (Y2) created by regression analysis as shown in Eq. 5.

$$\% \text{ Yield (Y2)} = 70.43 + 13.86 X_1 - 0.2025 X_2 - 0.6962 X_3 - 0.2425 X_1 X_2 + 1.86 X_1 X_3 - 1.43 X_2 X_3 + 6.47 X_1^2 - 6.75 X_2^2 - 3.83 X_3^2 \quad \text{Eq. (5)}$$

Equation 5 depicted that X₁ produced synergistic effect on percentage yield while X₂ and X₃ caused antagonistic effect on % yield of phytosomes. X₁, X₁², X₂² and X₃² have p -value less than 0.05 which showed their significant effect on Y2 (Table 3).^{21–23} The percentage yield rapidly increased with increase in lipid: extract due to presence of greater amount of lipids (Figure 5 and 6).^{39–41}

**Figure 5.** Illustration of effect of independent variables over percentage yield using two-dimensional contour plots

Optimization and validation of extract-loaded phytosomes

The desirability function (D-value) was investigated by Design-Expert to investigate optimized phytosome with pre-determined constraints of maximizing the percentage entrapment efficiency and percentage yield. The factors values for optimized phytosome were lipid: extract (X₁ = 1: 1 w/w), temperature (X₂ = 43°C) and time (X₃ = 2.15 hours). D-value for optimized phytosomes was found 0.929 having predicted Y1 and Y2 of 88.43% and 87.95%, respectively (Figure 7). Ramp plots (Figure 8) and overlay plot (Figure 9) demonstrated optimized values of factors and response variables of optimized phytosome.^{32,42,43} Actual values of Y1 and

Y2 for check point batch were found 87.32% and 85.87%, respectively. The percentage bias for Y1 and Y2 were 1.25% and 2.36%, respectively which validated the design model.

Table 3. Analysis of variance of percentage yield

Source	Sum of Squares	Df	Mean Square	F-value	p-value
Model	1967.50	9	218.61	80.55	< 0.0001
X ₁	1537.07	1	1537.07	566.37	< 0.0001
X ₂	0.3280	1	0.3280	0.1209	0.7422
X ₃	3.88	1	3.88	1.43	0.2855
X ₁ X ₂	0.2352	1	0.2352	0.0867	0.7803
X ₁ X ₃	13.76	1	13.76	5.07	0.0741
X ₂ X ₃	8.15	1	8.15	3.00	0.1436
X ₁ ²	154.68	1	154.68	57.00	0.0006
X ₂ ²	168.23	1	168.23	61.99	0.0005
X ₃ ²	54.23	1	54.23	19.98	0.0066
Lack of fit	6.13	3	2.04	0.5490	0.6965

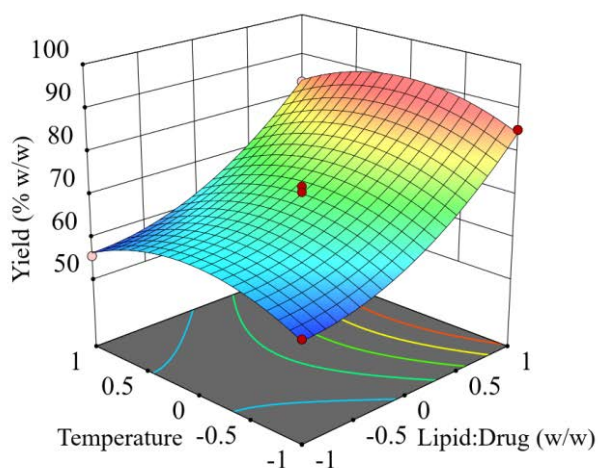
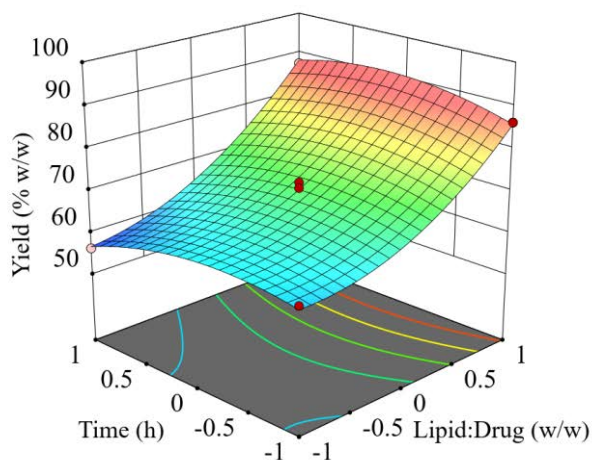


Figure 6. Depiction of effect of independent variables over percentage yield using three-dimensional response surface plots

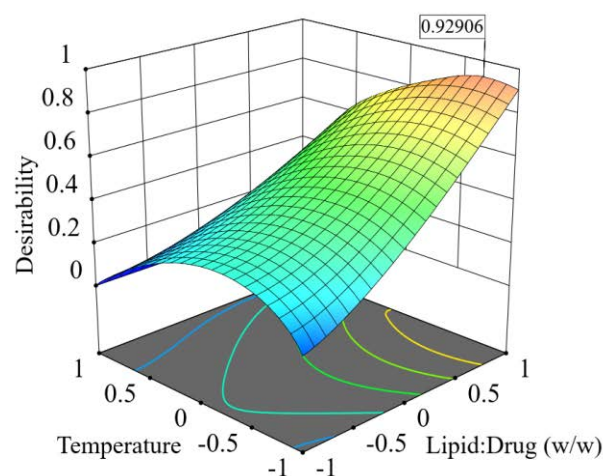
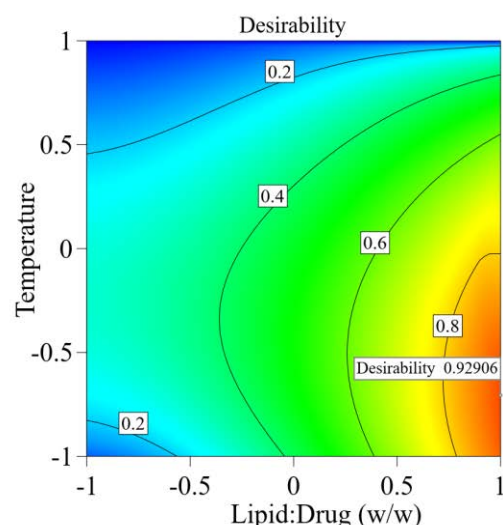


Figure 7. Contour and response surface plots depicting desirability function of optimized phytosome

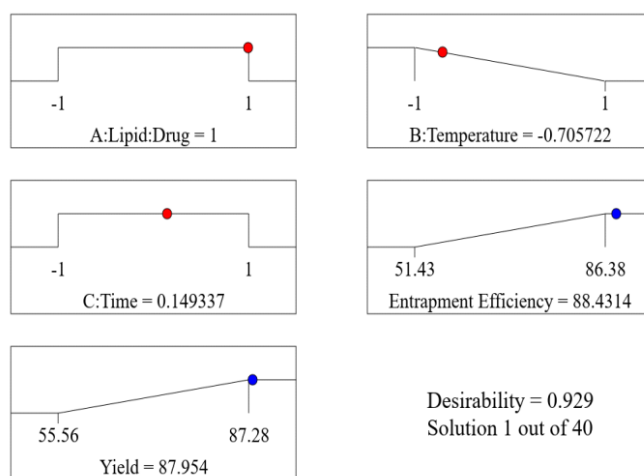


Figure 8. Ramp plots depicting optimized values of independent and dependent variables and desirability values of optimized phytosome

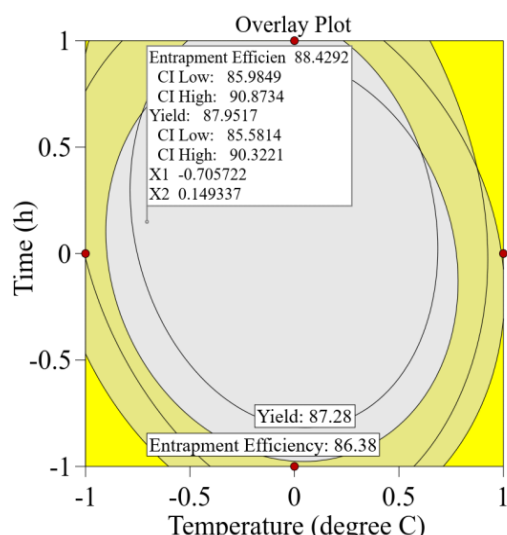


Figure 9. Overlay plot depicting predicted optimized values of independent and dependent variables (in square box) of optimized phytosome

CONCLUSIONS

In this research, *Andrographis paniculata* extracts loaded phytosomes were successfully synthesized by thin-film hydration procedure using soy lecithin and cholesterol. Box-Behnken design was used as optimization tool to explore optimized phytosomes. The research concluded that values of factors for optimized phytosomes were lipid: extract (X1 = 1: 1 w/w), temperature (X2 = 43°C) and time (X3 = 2.15 hours) having D-value 0.929. Furthermore, the percentage bias between actual and predicted values of Y1 and Y2 was 1.25% and 2.36%, respectively which concluded authenticity of design model.

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CONFLICTS OF INTEREST

The authors do not have any conflicts of interest.

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